

METABOLIC PRODUCTS OF
MICROORGANISMS 268[†]

OBSCUROLIDES, A NOVEL CLASS OF
PHOSPHODIESTERASE INHIBITORS
FROM STREPTOMYCES

II. MINOR COMPONENTS BELONGING
TO THE OBSCUROLIDE B TO D SERIES

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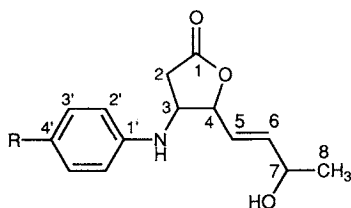
We previously²⁾ reported the isolation, structural
elucidation and biological properties of new phos-

phodiesterase inhibitors, the obscurolides A₁ to
A₄ (1 to 4) from the cultures of *Streptomyces*
viridochromogenes (strain Tü 2580). A careful search
for minor components by chemical screening
methods^{3,4)} revealed eight further members of the
obscurolide-complex²⁾ (Scheme 1). They were
assigned to the B, C and D series signifying their
differences in the polyketide moiety (C-1 to C-8),
whereas the subscripts indicated the oxidation level
of the functional group at C-4'. In this paper the
isolation, physico-chemical and biological char-
acterization of these new compounds are described.
All obscurolides were isolated as diastereomeric
mixtures at C-7 (A and C series) and C-5 (B series),
respectively. Only in the cases of 5/6 and 9/10
could the isomers be separated.

The raw product of a 10-liter fermentation²⁾ was
purified by flash chromatography on silica gel
(column 40 × 5 cm, CHCl₃-MeOH, 9:1) to give
1.9 g of the obscurolide-complex, which consisted
mainly of 2 and 3. The minor components were
isolated directly from the complex by preparative

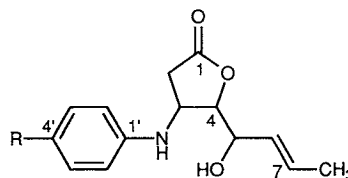
Scheme 1. Structures of the obscurolides belonging to the A to D series.

A series



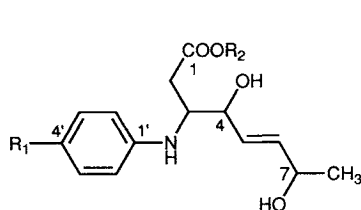
- 1 (A₁) R = COOH
2 (A₂) R = CHO
3 (A₃) R = CH₂OH
4 (A₄) R = CH₂OCH₃

B series



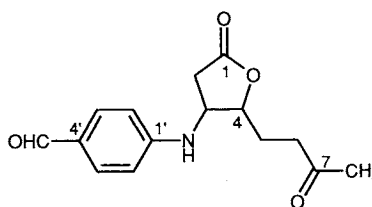
- 5 (B_{2α}) R = CHO
6 (B_{2β}) R = CHO
7 (B₃) R = CH₂OH
8 (B₄) R = CH₂OCH₃

C series



- 9 (C_{2α}) R₁ = CHO R₂ = H
10 (C_{2β}) R₁ = CHO R₂ = H
11 R₁ = CHO R₂ = CH₃

D series



- 12 (D₂)

[†] See ref 1).

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Table 1. Physico-chemical properties of the obscurolides B_{2α} to D₂ (5 to 12).

Compound	Rf-values ^a	Molecular formula	HREI-MS (M ⁺) found	Calcd for the given formula	IR (KBr cm ⁻¹)
B _{2α} (5)	I: 0.69 II: 0.30	C ₁₅ H ₁₇ NO ₄	275.1157	275.1152	3350, 1760, 1660
B _{2β} (6)	I: 0.78 II: 0.36	C ₁₅ H ₁₇ NO ₄	275.1154	275.1152	3340, 1770, 1655
B ₃ (7)	I: 0.38 II: 0.15	C ₁₅ H ₁₉ NO ₄	277.1314	277.1308	3470, 1770, 1705
B ₄ (8)	I: 0.60 II: 0.45	C ₁₆ H ₂₁ NO ₄	291.1471	291.1464	3380, 1770, 1710
C _{2α} (9)	I: 0.10 II: 0.04	C ₁₅ H ₁₉ NO ₅	^b	293.1257	3400, 1655, 1595
C _{2β} (10)	I: 0.10 II: 0.03	C ₁₅ H ₁₉ NO ₅	^b	293.1257	3400, 1655, 1600
11	I: 0.37 II: 0.07	C ₁₆ H ₂₁ NO ₅	307.1419	307.1413	3470, 1720, 1665
D ₂ (12)	I: 0.51 II: 0.21	C ₁₅ H ₁₇ NO ₄	275.1149	275.1152	3450, 1770, 1705

^a TLC silica gel, I: CHCl₃-MeOH (9:1), II: EtOAc-petroleum ether (1:1).

^b No EI-MS was obtainable.

Table 2. ¹H NMR signals in acetone-*d*₆ (200 MHz, δ in ppm relative to internal TMS) of the obscurolides B_{2α} to D₂ (5 to 12).

Proton	B _{2α} (5)	B _{2β} (6)	B ₃ (7) ^a	B ₄ (8) ^b	C _{2α} (9)	C _{2β} (10)	11 ^c	D ₂ (12)
2-Ha	2.44 dd	2.39 dd	2.35 dd	2.38 dd	2.40 dd	2.40 dd	2.62 dd	2.48 dd
2-Hb	3.13 dd	3.17 dd	3.06 dd	3.10 dd	2.55 dd	2.58 dd	2.76 dd	3.10 dd
3-H	4.43 m	4.42 m	4.32 m	4.38 m	4.24 m	4.21 m	4.38 m	4.06 m
3-NH	6.48 br d	6.49 br d	5.39 br d	5.51 br d	6.62 br d	6.60 br d	6.12 br d	6.46 br d
4-H	4.43 m	4.42 m	4.32 m	4.38 m	4.05 m	4.04 m	4.08 m	4.42 dt
5-H	4.43 m	4.42 m	4.32 m	4.38 m	5.79 m	5.80 m	5.77 m	1.85 ddt ^d
5-OH	4.56 d	4.75 d	2.93 br	2.90 br	3.03 br ^e	3.00 br ^e	4.52 br	2.21 m ^f
6-H	5.62 dd	5.64 dd	5.61 dd	5.66 dd	5.79 m	5.80 m	5.96 m	2.72 dd ^g
7-H	5.92 dq	5.93 dq	5.80 dq	5.85 dq	4.24 m	4.21 m	4.38 m	—
8-CH ₃	1.69* d	1.74* d	1.69 d	1.70 d	1.18 d	1.13 d	1.27* d	2.19 s
2'-H/6'-H	6.83 d	6.81 d	6.66 d	6.73 d	6.76 d	6.75 d	6.73 d	6.63 d
3'-H/5'-H	7.70 d	7.69 d	7.14 d	7.17 d	7.63 d	7.62 d	7.74 d	7.76 d
7'-H	9.73 s	9.74 s	4.47 s ^h	4.31 s ^h	9.73 s	9.73 s	9.77 s	9.79 s

^a 7'-OH: δ = 3.03 brs.

^b 8'-CH₃: δ = 3.29 q.

^c OCH₃: δ = 3.70 q.

^d 5-Ha.

^e 7-OH.

^f 5-Hb.

^g 6-H₂.

^h 7'-CH₂.

* Split signal, main peak.

reversed phase HPLC (Nucleosil 100 C-18, 10 μm, H₂O-MeOH, 70:30 to 0:100) in amounts of 9 mg (5), 7 mg (6), 9 mg (10), 8 mg (11) and 5 mg (12) and from fractions, which were obtained from the complex after flash chromatography on silica gel (column 40 × 5 cm, ethyl acetate-petroleum ether, 1:1) and subsequent chromatography on Sephadex

LH-20 with methanol yielding 25 mg of 7, 18 mg of 8 and 16 mg of 9. All obscurolides were easily detectable due to their UV-absorption at 254 nm and their yellow color on TLC plates after staining with EHRlich's reagent. Some physico-chemical properties of the new obscurolides are given in Table 1, whereas the ¹H and ¹³C NMR data are shown

Table 3. ^{13}C NMR signals (50.3 MHz, δ in ppm relative to internal TMS) of the obscurolides $\text{B}_{2\alpha}$ to D_2 (**5** to **12**) in acetone- d_6 , multiplicity assignments by attached proton test (APT).

Carbon	$\text{B}_{2\alpha}$ (5)	$\text{B}_{2\beta}$ (6)	B_3 (7)	B_4 (8) ^a	$\text{C}_{2\alpha}$ (9)	$\text{C}_{2\beta}$ (10)	11 ^b	D_2 (12)
1	175.7 s	175.5 s	176.3 s	176.2 s	179.9 s	180.0 s	172.8 s	174.8 s
2	36.5 t*	36.7 t*	36.8 t*	36.7 t*	39.7 t	40.4 t	35.3 t*	35.8 t
3	51.7 d*	50.1 d*	52.2 d*	52.1 d*	56.4 d	56.8 d	55.2 d	54.7 d
4	88.4 d	88.3 d	88.7 d	88.6 d	74.4 d	75.2 d	73.1 d*	84.8 d
5	73.2 d*	72.7 d*	73.4 d*	73.3 d	130.4 d	131.2 d	129.2 d*	28.3 t
6	129.1 d	128.9 d	128.7 d	128.7 d	137.2 d	138.0 d	137.6 d*	39.3 t
7	130.9 d	130.6 d	131.2 d	131.2 d	68.8 d	69.7 d	67.8 d*	206.5 s
8	17.9 q	18.0 q	17.9 q	17.9 q	23.4 q	24.0 q	23.9 q*	30.5 q
1'	153.3 s	153.0 s	147.0 s	148.8 s	155.8 s	156.4 s	154.2 s	153.3 s
2'/6'	113.2 d	113.2 d	113.9 d	113.8 d	113.5 d	114.0 d	113.2 d	113.2 d
3'/5'	132.7 d	132.6 d	129.1 d	130.2 d	133.6 d	134.2 d	132.5 d	132.6 d
4'	128.0 s	127.9 s	132.3 s	128.3 s	126.5 s	127.1 s	127.2 s	128.1 s
7'	190.2 d	190.2 d	64.7 t*	75.0 t	192.1 d	192.7 d	189.9 d	190.2 d

^a OCH_3 : $\delta = 57.5$ q.

^b OCH_3 : $\delta = 51.7$ q.

* Split signal, main peak.

in Table 2 and Table 3, respectively.

The molecular formula $\text{C}_{15}\text{H}_{17}\text{NO}_4$ of the obscurolides $\text{B}_{2\alpha}$ (**5**) and $\text{B}_{2\beta}$ (**6**) was identical with that of **2**, and the physico-chemical properties apart from the specific rotation ($[\alpha]_{\text{D}}^{20} = +65.9^\circ$ for **5** and $+23.4^\circ$ for **6**) were very similar. In the ^1H NMR spectra a downfield shift of 8- CH_3 from δ 1.22 (**2**) to δ 1.69 (**5**) and δ 1.74 (**6**) revealed the location of the double bond between C-6 and C-7. Accordingly 5-H showed an upfield shift from δ 5.95 (**2**) to δ 4.43/4.42 (**5/6**) demonstrating the shifting of the secondary hydroxy group from C-7 to C-5. Thus **5** and **6** (Scheme 1) differ in the stereochemistry at C-5 and for the substituents at C-3/C-4 we assume *trans* configuration as shown for an obscurolide A_2 derivative²). The related obscurolides B_3 (**7**) and B_4 (**8**) exhibited the same rearrangement of the allylic alcohol in the side chain as **5** and **6**. They were isomers of obscurolide A_3 (**3**) and A_4 (**4**), respectively, showing the typical signals for a hydroxymethylene group (**7**, δ_{H} 4.47, δ_{C} 64.7) and its corresponding methyl ether (**8**, δ_{H} 3.29/4.31, δ_{C} 57.5/75.0) attached to the benzene ring at C-4'.

The C series of the obscurolides combines those compounds, in which the γ -lactone was opened to the γ -hydroxycarboxylic acid (**9/10**) and its methyl ester (**11**), while the structure of the side chain follows the A series. Characteristic for all these compounds was the smaller distance of the ^1H NMR signals of the diastereotopic methylene protons 2-Ha and 2-Hb (e.g. δ 2.40 and 2.55 for **9**), which were fixed no longer in a γ -lactone ring. All open chain obscurolides had an aldehyde group in 4'-position,

Table 4. Effect of the obscurolides $\text{B}_{2\alpha}$ to D_2 (**5** to **12**) on calcium/calmodulin-dependent phosphodiesterase from bovine brain.

Obscurolide	IC_{50} (mM)
$\text{B}_{2\alpha}$ (5)	2.5
$\text{B}_{2\beta}$ (6)	> 15
B_3 (7)	1.5
B_4 (8)	0.4
$\text{C}_{2\alpha}$ (9)	10
$\text{C}_{2\beta}$ (10)	12
11	0.8
D_2 (12)	15

9 and **10** were diastereomers regarding C-7.

The only representative of the D series, obscurolide D_2 (**12**), showed a saturated side chain, in which starting from **2** the hydroxy group at C-7 has been oxidized to the ketone (δ_{C} 206.5).

The minor components of the obscurolide complex were tested as inhibitors of the calcium/calmodulin-dependent cAMP phosphodiesterase from bovine brain (Table 4)²). Although no definitive structure-activity relationship could be made with this limited data, it seems likely that the biological activity increases with the degree of reduction of the substituent at C-4'. The influence of the stereochemistry at C-5 becomes apparent if one compares the IC_{50} values of the diastereomers **5** and **6**. Although some of the minor components of the obscurolide complex are twenty-fold more active than the main components **2** and **3**, additional improvement was performed through enzymatic bromination, which will be the subject of a following

paper⁵). In analogy to the obscurolides of the A series the obscurolides of the B to D series revealed no growth inhibiting potency against bacteria, yeast and fungi.

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